The effect of smoking on caries-related microorganisms

Jiayi Wu¹, Mingyun Li^{2#}, Ruijie Huang^{2,3#}

ABSTRACT

INTRODUCTION Epidemiological studies have shown a close relationship between smoking and dental caries. Bacteria are one of the essential factors of caries formation. The imbalance of cariogenic bacteria and commensal bacteria in dental plaque results in higher production of acid that can corrode dental hard tissue. The aim of our review is to summarize the effect of smoking on caries-related bacteria.

METHODS English articles available in Pubmed and ScienceDirect databases and published before December 2018 were searched. A variety of evidence was collected including not only the influence of cigarette products on bacteria strains *in vitro* but also their effect on bacterial composition in saliva and dental plaque *in vivo*. We particularly emphasize the mechanisms by which nicotine acts on oral bacteria.

RESULTS The components of cigarettes promote the growth of cariogenic microorganisms. The mechanisms of how nicotine enhances *Streptococcus mutans, Lactobacilli, Streptococcus gordonii, Actinomyces* and *Candida albicans* are described separately in detail. The commensal bacteria, *Streptococcus sanguinis,* show less competitive capability in the presence of nicotine. Smoking influences saliva by lowering the buffer capability, altering its chemical agent and bacterial components, and therefore promotes the formation of a caries-susceptible environment. **CONCLUSIONS** Cigarette smoking and nicotine exposure promote the cariogenic activity of oral microorganisms and the formation of a caries-susceptible environment. This suggests that smokers should quit smoking, amongst other health reasons, also for their oral health.

AFFILIATION

 Department of Endodontic Dentistry, State Key Laboratory of Oral Diseases, National Clinical Research Center for Oral Diseases, West China Hospital of Stomatology, Sichuan University, Chengdu, China
State Key Laboratory of Oral Diseases, National Clinical Research Center for Oral Diseases, West China Hospital of Stomatology, Sichuan University, Chengdu, China

3 Department of Pediatric Dentistry, West China Hospital of Stomatology, Sichuan University, Chengdu, China "Contributed equally, co-correspondence authors

CORRESPONDENCE TO

Mingyun Li. State Key Laboratory of Oral Diseases, National Clinical Research Center for Oral Diseases, West China Hospital of Stomatology, Sichuan University, Chengdu, China. E-mail: limingyun@scu.edu.cn

Ruijie Huang. Department of Pediatric Dentistry, State Key Laboratory of Oral Diseases, National Clinical Research Center for Oral Diseases, West China Hospital of Stomatology, Sichuan University, Chengdu, China. E-mail: ruijmhuang@gmail.com ORCID ID: https://orcid.org/0000-0003-3211-518X

KEYWORDS

smoking, dental caries, nicotine, bacterial virulence factors

Received: 17 December 2018 Revised: 23 March 2019 Accepted: 24 March 2019

https://doi.org/10.18332/tid/105913

Tob. Induc. Dis. 2019;17(April):32

INTRODUCTION

Numerous epidemiological studies around the world have reported a close relationship between smoking and the occurrence of dental caries. In Italy, smoking military personnel (including 94.6% men and 5.4% women) have a higher decayed, missing, filled teeth (DMFT) score than non-smoking personnel¹. In Finland, daily smoking has been found to increase 4-year caries experience in adults². A study in Scotland has shown that if a pregnant woman smokes it might result in her child having a higher prevalence of caries than a child born to a non-smoking mother³. In Portugal, smoking has been confirmed as a risk factor for dental caries, and avoiding exposure to

Published by European Publishing on behalf of the International Society for the Prevention of Tobacco Induced Diseases (ISPTID). © 2019 Wu J. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 International License. (https://creativecommons.org/licenses/by/4.0/) smoking leads to a 7% decrease in caries incidence⁴. A systematic review by Benedetti et al.⁵ also verified that tobacco smoking had a close association with an increased risk of caries. However, there were differences between smokers and non-smokers in regard to personal education and economic situation⁶. Investigation showed that smokers tended to have bad eating habits, payed less attention to oral self-care, rarely seeking professional medical treatment and had poor compliance after treatment⁷⁻⁹. All these behaviors⁶⁻⁹ mentioned above could increase the incidence of caries. Further evidence to verify the cariogenic mechanisms of smoking needs to be researched. Nowadays, researchers have found that smoking has an effect on caries-related bacteria.

METHODS

Articles in English that were available in the Pubmed and ScienceDirect databases and published before December 2018 were searched. We collected and summarized the evidence of smoking and cigarette products not only as to how they influence the growth and metabolism of caries-related bacteria *in vitro* but also how they affect the saliva and dental plaque *in vivo*.

RESULTS

The components of cigarettes promote the growth of cariogenic microorganisms. The mechanisms of how nicotine enhances *Streptococcus mutans, Lactobacilli, Streptococcus gordonii, Actinomyces* and *Candida albicans* are described separately in detail. The commensal bacteria, *Streptococcus sanguinis,* show less competitive capability in the presence of nicotine. Smoking influences saliva by lowering the buffer capability, altering its chemical agent and bacterial components, therefore promoting the formation of a caries-susceptible environment.

The influence of cigarette products on the growth of oral bacteria

It is well known that bacteria create the preconditions for caries. Bacteria in the oral cavity produce acid by degrading the fermentable carbohydrates through the secretion of enzymes or metabolism, so as to induce further demineralization of dental hard tissues¹⁰. An early study¹¹ in 1991 claimed that smoking inhibited the growth of gram-positive cocci including *Neisseria*, one of the early colonizers in dental plaque. Smokers were considered to be inclined to form gram-negative bacterial colonization in their oral cavity, which was contradicted by later research. Baboni et al.¹² found that cigarette smoke condensate promoted the adhesion of Streptococcus mutans and Candida albicans to the acquired pellicle on orthodontic materials. Zonuz et al.¹³ incubated S. mutans and Streptococcus sanguis (now known as Streptococcus sanguinis) in atmospheric air, carbon dioxide and cigarette smoke. They found that cigarette smoke enhanced the growth of both bacteria strains, but S. mutans was affected more. One possible reason was that the former study did not examine the main cariogenic bacteria such as S. mutans. Another reason might be that there were differences in the experimental conditions, for example, different bacteria strains, cigarette varieties, different exposure time and so on. In addition, in vitro culture could not completely mimic the growth of bacteria in vivo.

There are about 7000 different kinds of molecules inhaled from smoking cigarettes¹⁴, making it difficult to ascertain the component that has the greatest effect on caries-related bacteria. More than 30 years ago, researchers believed it was the sugar content in tobacco that supported and influenced the growth of *S. mutans* and *S. sanguinis*, while chemical components including nicotine in tobacco did not affect oral microflora¹⁵. Nowadays, more and more evidence indicates that nicotine, the main bioactive and addictive substance in cigarette products, has a major impact on caries-related bacteria (summarized in Figure 1).

Streptococcus mutans and nicotine

S. mutans has been identified as the major pathogen of dental caries for its strong acid-resistant, acidogenic and biofilm forming abilities¹⁶. Compared to clinically healthy sites, the proportion of *S. mutans* is higher at white-spot enamel lesions¹⁷. In teeth with cavities, *S. mutans* even accounts for approximately 30% of the total bacteria, indicating that the proportion of *S. mutans* is related to the progressive stages of caries¹⁶.

Nicotine promotes the biofilm formation and metabolism of *S. mutans*¹⁸. One of our previous studies tested seven commonly used *S. mutans* strains: UA159, UA130, 10449, A32-2, NG8, LM7 and OMZ175¹⁸. For sub-minimum inhibitory

cariogenicity	status	MIC	MBC	MBIC	total growth	planktonic growth	biofilm formation	gene expression
S. mutans	main	16 ^[18]	32 ^[18]	8 or 16 ^[18]	↑ ^[13,20]	↓ ^[18]	↑ ^[18]	[19]
L. casei	pathogen	16 ^[34]	-	16 ^[34]	↑ ^[34]	~[34]	↑ ^[34]	-
C. albicans	conditioned pathogen	8 ^[56] or 16 ^[34]	-	16 ^[34]	↑ ^[34]	~[34]	↓ ^[56] or ↑ ^[34]	-
A. viscosus		16 ^[34]	-	16 ^[34]	↑ ^[34]	~[34]	↑ ^[34]	-
A. naeslundii		16 ^[34]	-	16 ^[34]	↓ ^[34]	~[34]	↓ ^[34]	-
S. gordonii		-	-	-	↑ ^[40]	1 ^[40]	↑ ^[40]	[40]
S.sanguinis	benign	-	-	-	↑ ^[13] or ↓ ^[20]	-	-	-

	function		mRNA level				protein level	
gene			F	ъс		вс	PC	BC
gtfB	surcrose-			Î		↓*	-	-
gtfC	dependent E synthesis and			1		↓*	-	-
gtfD	bindings	CON		↑ ↓*		↓*	-	-
gbpA				î		~	↑*	~
gbpB	biofilm format	tion		î		↓*	~	↓*
gbpC	and virulend	e		î		~	-	-
gbpD				î	↓*		-	-
		_	_		_			
gene	function	P	C gene		function		PC	
abpA	cell-binding and biofilm formation	1	*	sspB				~
abpB		~		hsa			-binding	~
ссрА		^*		cshA		and biofilm formation		~
srtA		1	ł	cshB				~
gtfG		~		sca	A			† *
sspA		~			mRNA level			

A green to red color gradient indicates an increase in cariogenicity. The unit of MIC/MBC/MBIC is mg/mL. The influence of nicotine on total growth, planktonic growth and biofilm formation are all estimated at sub-MIC level. PC: planktonic cells, BC: biofilm cells. \uparrow represents facilitation. ~ represents no significant effects. \downarrow represents inhibition. Absence of * next to gene expression indicates that only trends are not statistically significant

concentration (sub-MIC: nicotine concentrations 0, 0.25, 0.5, 1, 2, 4 and 8 mg/mL), the majority of S. mutans strains' planktonic growth was not affected at low nicotine concentrations (0.25-2 mg/mL) but was inhibited at 4 and 8 mg/mL. In human saliva, the nicotine concentration is dependent on several factors such as salivary flow rate, daily cigarettes smoked, the lapse after last cigarette, individual differences etc¹⁸. It seemed that at human saliva nicotine concentration range, nicotine had no effect on S. mutans planktonic growth. However, the S. mutans in biofilm was critical for caries development but not in saliva. For the same seven species, bacterial biofilm formation was significantly increased even at 0.5 mg/mL nicotine¹⁸. This result was consistent with the work of Zonuz et al.¹³. For UA159, a strain of S. mutans, the biofilm mass was clearly shown in scanning electron microscopy (SEM). In nicotine treated groups, more biofilm was formed composed of bacterial cells and extracellular polymeric substances (EPS). The biofilm was more structurally formed with longer bacterial chain length and more orientated cell arrangement than the control¹⁸. To better illustrate and guantify the augmentation of bacterial cells and EPS separately, confocal laser scanning microscopy (CLSM) has been used. Both bacterial cells and EPS were significantly increased at 2 and 4 mg/mL nicotine^{19,20}.

The question is whether more bacteria cells result in more active metabolism and more acid produced. A 2,3-bis(2-methoxy-4-nitro-5-sulfophenyl)-2Htetrazolium-5-carboxanilide (XTT) assay, which can test the general bacteria metabolism, indicated increased overall bacterial metabolic activity for all of the seven *S. mutans* species¹⁸. To calculate the net metabolism increment of a unit cell, the overall metabolism was divided by the biofilm mass. Data indicated that even for a unit cell, the metabolism was increased by nicotine¹⁸. Lactate dehydrogenase (LDH) is the enzyme catalyzing the conversion of pyruvate to lactate and producing lactic acid as final product²¹. The overall LDH activity of *S. mutans* biofilm is increased by nicotine, but for a unit cell the LDH activity is not changed¹⁹.

EPS is made of polysaccharides, proteins, eDNA, lipid and other macromolecules. It is essential for the structural and functional integrity of dental biofilm²². Glucosyltransferases (Gtfs) are special enzymes that are critically involved in carbohydrate degradation, sucrose-dependent EPS synthesis and cell bindings of S. mutans. Gtfs degrade sucrose into glucose and fructose firstly and then utilize glucose to synthesize glucan²³. S. mutans can express three distinct types: *gtfB*, *gtfC* and *gtfD*. They are responsible for a-1,3-rich water-insoluble glucan, a-1,3-rich water-insoluble glucan and certain a-1,6-rich water-soluble glucan, and a-1,6-rich water-soluble glucan, respectively²⁴⁻²⁶. The gene expression of gtfB, gtfC and gtfD of planktonic S. mutans cells indicated an increased trend at 2 mg/mL nicotine compared with the control group, but the increment was not statistically significant due to a large standard deviation. On the other hand, the expression of those three genes of

S. mutans cells in biofilm was decreased. Those *gtfs* results seem contradictory to previous increased biofilm data¹⁹. One possible hypothesis is that as cells increase in the biofilm, bacteria repel more cell attachment to protect their own nutrients through a Quorum sensing (QS) system. As for the planktonic cells, the effect of QS system is not as strong as that in biofilm. Further experiments are needed before any conclusion can be drawn to explain those conflicting observations.

S. mutans can also express a series of glucan binding proteins (Gbps). The discovery of GbpA was in 1990 by Banas et al.²⁷, followed by GbpB in 1994 by Smith et al.²⁸, GbpC in 1997 by Sato et al.²⁹ and GbpD in 2004 by Shah and Russell³⁰. Those four Gbps are associated with the biofilm formation and virulence of S. mutans, but the functions of Gbps have not been systematically analyzed³¹. The expressions of GbpA and GbpB are higher in planktonic cells than in biofilm cells¹⁹, which might indicate that the bacteria in planktonic status are more vigorously seeking a place to bind to than those in biofilm status. At protein level for planktonic S. mutans cells, nicotine significantly promotes the expression of GbpA but has no effect on GbpB, while for biofilm cells, nicotine has no effect on GbpA but significantly inhibits the expression of GbpB¹⁹. At mRNA level, nicotine indicates a non-significant promoting effect on gbpA, gbpB, gbpC and gbpD expression of planktonic S. mutans cells but a significant inhibitory effect on gbpB and gbpD expression of biofilm cells¹⁹.

Lactobacilli and nicotine

Lactobacilli are another chief pathogen associated with caries. Ruyven et al.³² detected Lactobacilli rather than non-mutans streptococci and Actinomyces from dental biofilms at white-spot lesions. Other studies found that Lactobacilli were more prevalent than S. mutans at the advancing front of dentin caries¹⁶. Like S. mutans, Lactobacilli are also acidogenic and aciduric. L. casei as well as other Lactobacilli can produce a significant amount of lactic acid and can remain viable under severely acidic conditions³³.

Both the MIC and minimum biofilm inhibitory concentration (MBIC) of nicotine on *L. casei* are 16 mg/mL³⁴. The planktonic growth of *L. casei* is not significantly affected at sub-MIC level. Nicotine can induce a distinct upward trend in the biofilm

formation of *L. casei*. An increasing trend can be seen when considering the planktonic growth and the biofilm formation as a whole. Detailed mechanisms about how nicotine moderates the growth of *L. casei* have not been clarified.

Streptococcus sanguinis and nicotine

S. sanguinis and S. mutans compete for the same ecological niche and have similar metabolic characters³⁵. S. sanguinis can produce hydrogen peroxide and sanguicin to inhibit the growth of S. mutans, moreover, it can also inhibit S. mutans bacteriocins synthesis. Clinical studies found that S. sanguinis was predominant over S. mutans in dental plaque of healthy individuals compared to those with caries³⁶⁻³⁸. As a result, S. sanguinis is usually considered benign for caries.

The growth of mono-cultured S. sanguinis and S. mutans increased as the ratio of nicotine and tar content increased¹³. However, nicotine only slightly promoted S. sanguinis but significantly enhanced S. mutans. Our study constructed a double-bacteria hybrid model and found that the ratio of S. mutans/S. sanguinis increased as nicotine concentration increased. In contrast to S. mutans, nicotine treatment had no effect on S. sanguinis colony forming unit (CFU). Fluorescence in situ hybridization (FISH) results confirmed that the ratio of S. mutans/S. sanguinis was significantly increased at 4 mg/mL nicotine after 24 hours treatment²⁰. As for the biofilm formation, the ratio was significantly increased at 48 and 72 hours at 1 mg/mL nicotine²⁰. In summary, higher nicotine concentration can boost the advantage of S. mutans by facilitating S. mutans to compete with S. sanguinis. This effect is less but still apparent at a lower nicotine concentration.

Streptococcus gordonii and nicotine

The role of *S. gordonii* in caries is controversial. On the one hand, it can produce hydrogen peroxide to inhibit *S. mutans* growth, even though the amount of hydrogen peroxide produced is less than that of *S. sanguinis*³⁷. But on the other hand, *S. gordonii* provides binding sites and facilitates the attachment of *S. mutans* to the tooth surface³⁹.

Nicotine (0.5–4 mg/mL) significantly promotes *S. gordonii* planktonic cell growth at all the tested time points: 12 hours, 24 hours and 48 hours⁴⁰.

The promotion effect is consistent among most varied concentration groups. The total biofilm mass is significantly increased at 0.5-4 mg/mL nicotine after 24 hours incubation⁴⁰. Analysis of the biofilm composition found that bacterial cell number is doubled in 2 mg/mL nicotine and 2.5-fold in 4 mg/ mL nicotine. Though the amount of EPS indicates an increased trend, it is non-significant due to a large standard deviation⁴⁰. For S. gordonii cell aggregation assay, in the presence of sucrose, the aggregation increases after nicotine concentration reaches 4 mg/mL. Without sucrose, only 16 mg/mL nicotine promotes the aggregation. These findings imply that sucrose is involved in the process of nicotine promoting cell aggregation. The combined action of sucrose and nicotine enhances the cell binding and cell attachment properties of S. gordonii.

The expression of eleven cell-binding and biofilmforming related genes of S. gordonii has been investigated. S. gordonii binds to amylase, a salivary enzyme that hydrolyzes starch to monosaccharide, via amylase-binding protein A (AbpA) and amylasebinding protein B (AbpB)⁴¹⁻⁴³. EPS synthesis and biofilm formation are controlled by carbon catabolite protein A (CcpA), which is directly related to sucrose metabolism⁴⁴. CcpA mutant strain can hardly generate biofilm. Glucosyltransferase G (*GtfG*) is involved in synthesizing water-soluble and water-insoluble glucans and in regulating S. gordonii adhesion⁴⁵. Sortase A (SrtA) regulates S. gordonii cell wall anchoring transpeptidase and peptidoglycan synthesis^{46,47}. Streptococcal surface protein A (SspA) and Streptococcal surface protein B (SspB) encode S. gordonii antigen I/II adhesion. Specially, SspA regulates salivary agglutinin aggregation^{48,49}. Streptococcal hemagglutinin (Hsa) involves in S. gordonii sialic acid binding⁵⁰. Surfaceassociated proteins CshA, CshB and substrate-binding lipoproteins (ScaA) also play a role in bacterial cell surface recognition and bindings^{45,51}. Among these eleven proteins, nicotine stimulates the gene expression of abpA, scaA, ccpA and srtA. Though not promoted, none of the remaining genes is inhibited by nicotine⁴⁰.

Actinomyces and nicotine

Non-mutans streptococci and Actinomyces are predominant in caries-free individuals, and they

play a role in the initiation of caries. Though both are acidogenic and aciduric, their abilities are not as strong as those of *S. mutans* and *lactobacilli*. Their final pH value can be lower than pH 5.5, which is the critical acid concentration where demineralization occurs¹⁶.

Aubrey et al.³⁴ estimated the effect of nicotine on Actinomyces viscosus and Actinomyces naeslundii, two common kinds of Actinomyces, and found that both MIC and minimum bactericidal concentration (MBC) were 16 mg/mL. However, A. viscosus and A. naeslundii showed an opposite trend in total growth and biofilm formation treated with double-diluted nicotine. The former had an increase in growth and biofilm formation by 8 mg/mL of nicotine, while the latter decreased by 8 mg/mL and was completely inhibited at 16 mg/mL of nicotine.

Candida albicans and nicotine

C. albicans is normally a ubiquitous harmless fungus in human oral cavity, upper respiratory tract, intestines and vagina. It is an opportunistic pathogen that often causes candidiasis in immunocompromised patients with HIV/AIDS for example^{52,53}. C. albicans is also a cariogenic microbe with abilities to adhere to tooth surfaces and produce acid⁵⁴. Investigations indicate that C. albicans has a close relationship with early childhood caries (ECC)⁵⁵. One experiment indicated that the MIC of nicotine on C. albicans was 8 mg/ mL56 and another that it was 16mg/mL³⁴. The latter³⁴ study also considered 16 mg/mL as MBIC. These two studies came to different conclusions about the biofilm formation of mono-cultural C. albicans treated with nicotine. The former⁵⁶ found that nicotine did not affect the biofilm formation at lower concentrations (1 and 2 mg/mL) but displayed an inhibitory effect at higher concentrations (4 mg/mL). Nevertheless, the latter³⁴ found that nicotine could dose-dependently promote the biofilm formation as well as the total growth of mono-cultural C. albicans through 8 mg/ mL. More research is needed to reconcile these differences.

Increased interests seem to focus on the interaction between *S. mutans* and *C. albicans* during the development of caries. *S. mutans* has been detected together with *C. albicans* in high numbers in dental plaque from ECC. In single-pathogen infected rats, the severity of caries lesions became worse when treated with dual-pathogens⁵⁷. Studies^{58,59} showed that *C. albicans* promoted the adherence of S. mutans. SEM analysis of dual-species biofilm indicated that *C. albicans* cells in the mixed biofilm were found to be surrounded by *S. mutans* cells. *S. mutans* exhibited high affinity to *C. albicans* hyphae. Nicotine slightly increased the biomass of the mixed biofilm. The proportion of *C. albicans* increased when treated with 1 and 2 mg/mL nicotine⁵⁶. However, only a few *C. albicans* were present in the dual-species biofilm when treated without nicotine.

The influence of smoking on oral bacterial compositions

Bacterial composition in saliva in vivo

Several studies⁶⁰⁻⁶² have tested the numbers of cariogenic bacteria of whole saliva collected from smokers and non-smokers. They came to the consistent conclusion that significantly higher numbers of *Lactobacilli* were observed in smokers. Though the counts of *S. mutans* also increased in smokers, the rising degrees were different. Heintze et al.⁶⁰ reported that *S. mutans* were significantly higher in saliva of smokers. Sakki et al.⁶¹ found that smoking induced an unapparent increase in *S. mutans*. They also observed significantly higher presence of yeasts that were nonpathogenic form of *C. albicans* in smokers' saliva⁶³.

Bacterial composition in dental plaque

Other than planktonic bacteria in saliva, dental plaque, which is the preferred living medium for bacteria, plays a key role in caries development. It provides a suitable environment for bacteria to grow, metabolize and reproduce, promotes bacteria to adhere to the tooth surface, allows interactions among different microorganisms and helps to resist external stimuli^{64,65}. A study⁶⁶ estimated the plaque index and wet weight of mature plaque, and found that smoking did not significantly promote the formation rate of dental plaque. As for chemical compositions, compared to non-smokers, the protein nitrogen, calcium or (total) phosphorus concentrations in dental plaque of smokers were not significantly different. However, Ca/P ratio was markedly higher in smokers' dental plaque. Considering the calcium concentration was higher only when relative to phosphorus concentration, the extra calcium might not play a part in mineral deposition but affect calculus formation.

The formation of dental plaque includes three main stages⁶⁷, and the effect of smoking on each stage is illustrated in Figure 2.

In the first stage, salivary proteins and glycoproteins stick to the tooth surface and form acquired pellicle, which protects the tooth against demineralization. The protective functions of acquired pellicle depend on its thickness, maturation and composition, which are influenced by the characteristics of saliva⁶⁸. Studies have found that smoking does not lower salivary flow or secretion rate⁶⁰ but lowers the buffer capability⁶⁹ of saliva, which results in a possible lower pH of saliva. It also influences the concentration of saliva proteins such as salivary secretory IgA (sIgA) and amylase, which can be stably detected in acquired pellicle. sIgA is considered the main defence agent against oral diseases by preventing microbial adherence to tooth surfaces and oral epithelial cells. Numerous studies have verified that lower concentration of sIgA is a risk factor for dental caries in both children and adults⁶⁹⁻⁷¹. Smoking has a dose-dependent immunosuppressive property, which is reflected by the decrease of sIgA concentration in saliva in both adult smokers and passive-smoking children. Amylase is crucial to the colonization and metabolism of streptococci, which contributes to the occurrence of caries. Amylase acts as a receptor in the acquired pellicle for bacteria to adhere to the tooth surface. Previous studies reported that passive smokers had a higher concentration of amylase in their saliva⁷¹.

In the second stage, also called initial adhesion stage, bacteria recognize the glycoproteins in the acquired pellicle and start to bind to them. The pioneer bacteria of this stage are almost all Streptococci, i.e. *Streptococcus mitis, S. sanguinis, Streptococcus oralis* and *S. gordonii*³⁹. The community of the biofilm is relatively simple and the Streptococci reach their highest count by percentage.

In the third stage, also called biofilm maturation stage, later colonizers recognize the adhesion receptors expressed on the cell surface of pioneer bacteria and start to reside in the biofilm, making a more complex community with numerous species. The count of Streptococci by percentage decreases (although the absolute number increases). The predominant bacteria shift from Streptococci solely to Streptococci and Actinomyces together¹⁶. Most of

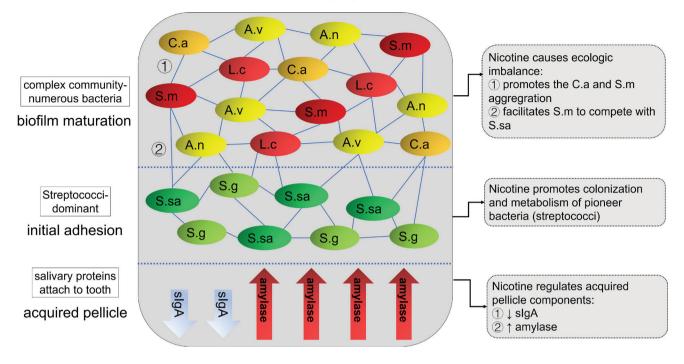


Figure 2. The effect of smoking on each stage of dental plaque formation

the Streptococci are still non-mutans with very low numbers of *S. mutans* on clinically sound enamel surfaces. Main cariogenic pathogens such as *S. mutans* and *Lactobacillus* also seek their home in the biofilm during this stage⁶⁷, and their number is higher in people with caries, which means changes in the components of bacteria in the biofilm are related to the degree of progression of caries¹⁶.

DISCUSSION

In current opinion, having caries is no longer regarded as a disease caused by one pathogen. The cariogenic bacteria may come from commensal oral flora. There are dynamic ecological balances between oral bacteria themselves as well as between bacteria and host. When the balances are compromised, these commensal bacteria become pathogens and thus lead to caries⁷². This etiological hypothesis is in agreement with the descriptions in the previous sections that almost all bacterial species play a certain role in caries. For this reason, taking all the bacteria in the dental plaque as a whole and finding the specific bacterial composition changes in dental plaque from smokers and non-smokers will be of more clinical significance. However, there is no such report up to now.

CONCLUSIONS

This review shows a close relationship between smoking and caries-related microorganisms. Though the effects of extended tobacco use may not be reversible⁷³, it is better to quit smoking as soon as possible. Besides, there are still questions on this topic, such as the effect of nicotine on other oral bacteria that have not been mentioned in this article, which need further investigation if one is to come to more robust conclusions.

REFERENCES

- Campus G, Cagetti MG, Senna A, Blasi G, Mascolo A, Demarchi P, Strohmenger L. Does smoking increase risk for caries? a cross-sectional study in an Italian military academy. Caries Res. 2011;45(1):40-46. doi:10.1159/000322852
- Bernabé E, Delgado-Angulo EK, Vehkalahti MM, Aromaa A, Suominen AL. Daily smoking and 4-year caries increment in Finnish adults. Community Dentistry & Oral Epidemiology. 2015;42(5):428-434. doi:10.1111/cdoe.12101
- Bernabé E, Macritchie H, Longbottom C, Pitts NB, Sabbah W. Birth Weight, Breastfeeding, Maternal Smoking and Caries Trajectories. J Dent Res. 2017;96(2):171-178. doi:10.1177/0022034516678181
- 4. Nobre MA, Malã P. Prevalence of periodontitis, dental caries, and peri-implant pathology and their relation

with systemic status and smoking habits: Results of an open-cohort study with 22009 patients in a private rehabilitation center. J Dent. 2017;67:36-42. doi:10.1016/j.jdent.2017.07.013

- Benedetti G, Campus G, Strohmenger L, Lingström P. Tobacco and dental caries: a systematic review. Acta Odontol Scand. 2013;71(3-4):363-371. doi:10.3109/00016357.2012.734409
- 6. Beaglehole R, Myriad Editions, International Dental Federation. The oral health atlas : mapping a neglected global health issue. Cointrin, Switzerland: FDI World Dental Federation; 2009.
- Mucci LA, Brooks DR. Lower use of dental services among long term cigarette smokers. J Epidemiol Community Health. 2001;55(6):389-393. doi:10.1136/jech.55.6.389
- Sherwood NE, Hennrikus DJ, Jeffery RW, Lando HA, Murray DM. Smokers with multiple behavioral risk factors: how are they different? Prev Med. 2000;31(4):299-307. doi:10.1006/pmed.2000.0710
- Axelsson P, Paulander J, Lindhe J. Relationship between smoking and dental status in 35-, 50-, 65-, and 75-yearold individuals. J Clin Periodontol. 1998;25(4):297-305. doi:10.1111/j.1600-051x.1998.tb02444.x
- Takahashi N, Nyvad B. Ecological Hypothesis of Dentin and Root Caries. Caries Res. 2016;50(4):422-431. doi:10.1159/000447309
- 11. Ertel A, Eng R, Smith SM. The differential effect of cigarette smoke on the growth of bacteria found in humans. Chest. 1991;100(3):628-630. doi:10.1378/chest.100.3.628
- Baboni FB, Guariza FO, Moreno AN, Rosa E. Influence of cigarette smoke condensate on cariogenic and candidal biofilm formation on orthodontic materials. Am J Orthod Dentofacial Orthop. 2010;138(4):427-434. doi:10.1016/j.ajodo.2009.05.023
- Zonuz AT, Rahmati A, Mortazavi H, Khashabi E, Farahani RM. Effect of cigarette smoke exposure on the growth of Streptococcus mutans and Streptococcus sanguis: an in vitro study. Nicotine Tob Res. 2008;10(1):63-67. doi:10.1080/14622200701705035
- 14. Courtney R. The Health Consequences of Smoking-50 Years of Progress: A Report of the Surgeon General, 2014Us Department of Health and Human Services Atlanta, GA: Department of Health and Human Services, Centers for Disease Control and Prevention, National Center for. Drug Alcohol Rev. 2015;34(6):694-695. doi:10.1111/dar.12309
- Lindemeyer RG, Baum RH, Hsu SC, Going RE. In vitro effect of tobacco on the growth of oral cariogenic streptococci. J Am Dent Assoc. 1981;103(5):719-722. doi:10.14219/jada.archive.1981.0372
- Takahashi N, Nyvad B. The role of bacteria in the caries process: ecological perspectives. J Dent Res. 2011;90(3):294-303. doi:10.1177/0022034510379602
- 17. Van HJ, Sansone C, Joshipura K, Kent R. Mutans

streptococci and non-mutans streptococci acidogenic at low pH, and in vitro acidogenic potential of dental plaque in two different areas of the human dentition. J Dent Res. 1991;70(12):1503-1507. doi:10.1177/00220345910700120601

- Huang R, Li M, Gregory RL. Effect of nicotine on growth and metabolism of Streptococcus mutans. Eur J Oral Sci. 2012;120(4):319-325. doi:10.1111/j.1600-0722.2012.00971.x
- Huang R, Li M, Gregory RL. Nicotine promotes Streptococcus mutans extracellular polysaccharide synthesis, cell aggregation and overall lactate dehydrogenase activity. Arch Oral Biol. 2015;60(8):1083-1090. doi:10.1016/j.archoralbio.2015.04.011
- 20. Li M, Huang R, Zhou X, Zhang K, Zheng X, Gregory RL. Effect of nicotine on dual-species biofilms of Streptococcus mutans and Streptococcus sanguinis. FEMS Microbiol Lett. 2014;350(2):125-132. doi:10.1111/1574-6968.12317
- 21. Sommer P, Klein JP, Sch Ller M, Frank RM. Lactate dehydrogenase from Streptococcus mutans: purification, characterization, and crossed antigenicity with lactate dehydrogenases from Lactobacillus casei, Actinomyces viscosus, and Streptococcus sanguis. Infect Immun. 1985;47(2):489-495. https://www.ncbi.nlm.nih.gov/ pubmed/3917978. Accessed December 17, 2018.
- Sutherland IW. The biofilm matrix an immobilized but dynamic microbial environment. Trends Microbiol. 2001;9(5):222-227. doi:10.1016/s0966-842x(01)02012-1
- Bowen WH, Koo H. Biology of Streptococcus mutans-Derived Glucosyltransferases: Role in Extracellular Matrix Formation of Cariogenic Biofilms. Caries Res. 2011;45(1):69-86. doi:10.1159/000324598
- 24. Aoki H, Shiroza T, Hayakawa M, Sato S, Kuramitsu HK. Cloning of a Streptococcus mutans glucosyltransferase gene coding for insoluble glucan synthesis. Infect Immun. 1986;53(3):587-594. PMID: 3017865.
- Hanada N, Kuramitsu HK. Isolation and characterization of the Streptococcus mutans gtfC gene, coding for synthesis of both soluble and insoluble glucans. Infect Immun. 1988;56(8):1999-2005. PMID: 2969375.
- Hanada N, Kuramitsu HK. Isolation and characterization of the Streptococcus mutans gtfD gene, coding for primerdependent soluble glucan synthesis. Infect Immun. 1989;57(7):2079-2085. PMID: 2543630.
- Banas JA, Russell RR, Ferretti JJ. Sequence analysis of the gene for the glucan-binding protein of Streptococcus mutans Ingbritt. Infect Immun. 1990;58(3):667-673. PMID: 2307516.
- Smith DJ, Akita H, King WF, Taubman MA. Purification and antigenicity of a novel glucan-binding protein of Streptococcus mutans. Infect Immun. 1994;62(6):2545-2552. PMID: 8188378.
- 29. Sato Y, Yamamoto Y, Kizaki H. Cloning and sequence analysis of the gbpC gene encoding a novel glucan-

binding protein of Streptococcus mutans. Infect Immun. 1997;65(2):668-675. PMID: 9009329.

- 30. Shah DS, Russell RR. A novel glucan-binding protein with lipase activity from the oral pathogen Streptococcus mutans. Microbiology. 2004;150(6):1947-1956. doi:10.1099/mic.0.26955-0
- 31. Lynch DJ, Fountain TL, Mazurkiewicz JE, Banas JA. Glucan-binding proteins are essential for shaping Streptococcus mutans biofilm architecture. Fems Microbiol Lett. 2010;268(2):158-165. doi:10.1111/j.1574-6968.2006.00576.x
- 32. van Ruyven FO, Lingsträ MP, Van HJ, Kent R. Relationship among mutans streptococci, "low-pH" bacteria, and lodophilic polysaccharide-producing bacteria in dental plaque and early enamel caries in humans. J Dent Res. 2000;79(2):778-784. doi:10.1177/00220345000790021201
- 33. Hujanen M, Linko S, Linko YY, Leisola M. Optimisation of media and cultivation conditions for L(+)(S)-lactic acid production by Lactobacillus casei NRRL B-441. Appl Microbiol Biotechnol. 2001;56(1-2):126-130. doi:10.1007/s002530000501
- 34. DuBois AE, Bennett ZC, Khalid U, Khalid A, Meece RA, Difiore GJ, Gregory RL. Nicotine: Its Stimulating and Inhibitory Effects on Oral Microorganisms. Fine Focus. 2014;1:63-75. http://cardinalscholar.bsu.edu/bitstream/ handle/123456789/201343/Fine%20Focus%20 1%281%29%20p63-75.pdf?sequence=1&cisAllowed=y. Accessed December 17, 2018.
- 35. Valdebenito B, Tullume-Vergara PO, González W, Kreth J, Giacaman RA. In silico analysis of the competition between Streptococcus sanguinis and Streptococcus mutans in the dental biofilm. Mol Oral Microbiol. 2017;33(2):168-180. doi:10.1111/omi.12209
- 36. Kreth J, Merritt J, Shi W, Qi F. Co-ordinated bacteriocin production and competence development: a possible mechanism for taking up DNA from neighbouring species. Mol Microbiol. 2010;57(2):392-404. doi:10.1111/j.1365-2958.2005.04695.x
- 37. Kreth J, Zhang Y, Herzberg MC. Streptococcal antagonism in oral biofilms: Streptococcus sanguinis and Streptococcus gordonii interference with Streptococcus mutans. J Bacteriol. 2008;190(13):4632-4640. doi:10.1128/jb.00276-08
- 38. Tong H, Chen W, Shi W, Fengxia QI, Dong X. SO-LAAO, a Novel L-Amino Acid Oxidase That Enables Streptococcus oligofermentans To Outcompete Streptococcus mutans by Generating H2O2 from Peptone. J Bacteriol. 2008;190(13):4716-4721. doi:10.1128/jb.00363-08
- Kolenbrander PE, Palmer RJ, Rickard AH, Jakubovics NS, Chalmers NI, Diaz PI. Bacterial interactions and successions during plaque development. Periodontol 2000. 2006;42(1):47-79. doi:10.1111/j.1600-0757.2006.00187.x
- 40. Huang R, Li M, Ye M, Yang K, Xu X, Gregory RL. Effects of Nicotine on Streptococcus gordonii Growth, Biofilm

Formation, and Cell Aggregation. Appl Environ Microbiol. 2014;80(23):7212-7218. doi:10.1128/aem.02395-14

- 41. Li L, Tanzer JM, Scannapieco FA. Identification and analysis of the amylase-binding protein B (AbpB) and gene (abpB) from Streptococcus gordonii. FEMS Microbiol Lett. 2002;212(2):151-157. doi:10.1111/j.1574-6968.2002.tb11259.x
- 42. Rogers JD, Haase EM, Brown AE, Douglas CW, Gwynn JP, Scannapieco FA. Identification and analysis of a gene (abpA) encoding a major amylase-binding protein in Streptococcus gordonii. Microbiology. 1998;144(5):1223-1233. doi:10.1099/00221287-144-5-1223
- Aguirre A, Levine MJ, Cohen RE, Tabak LA. Immunochemical quantitation of α-amylase and secretory IgA in parotid saliva from people of various ages. Arch Oral Biol. 1987;32(4):297-301. doi:10.1016/0003-9969(87)90024-0
- 44. Zheng L, Chen Z, Itzek A, Herzberg MC, Kreth J. CcpA regulates biofilm formation and competence in Streptococcus gordonii. Mol Oral Microbiol. 2012;27(2):83-94. doi:10.1111/j.2041-1014.2011.00633.x
- 45. Vickerman MM, Sulavik MC, Nowak JD, Gardner NM, Jones GW, Clewell DB. Nucleotide sequence analysis of the Streptococcus gordonii glucosyltransferase gene, gtfG. DNA Seq. 1997;7(2):83-95. doi:10.3109/10425179709020155
- Li MY, Huang RJ, Zhou XD, Gregory RL. Role of sortase in Streptococcus mutans under the effect of nicotine. Int J Oral Sci. 2013;5(4):206-211. doi:10.1038/ijos.2013.86
- Paterson GK, Mitchell TJ. The biology of Gram-positive sortase enzymes. Trends Microbiol. 2004;12(2):89-95. doi:10.1016/j.tim.2003.12.007
- Holmes AR, McNab R, Jenkinson HF. Candida albicans binding to the oral bacterium Streptococcus gordonii involves multiple adhesin-receptor interactions. Infect Immun. 1996;64(11):4680-4685. PMID: 8890225.
- 49. Jenkinson HF, Terry SD, McNab R, Tannock GW. Inactivation of the gene encoding surface protein SspA in Streptococcus gordonii DL1 affects cell interactions with human salivary agglutinin and oral actinomyces. Infect Immun. 1993;61(8):3199-3208. PMID: 8335350.
- 50. Takahashi Y, Konishi K, Cisar JO, Yoshikawa M. Identification and characterization of hsa, the gene encoding the sialic acid-binding adhesin of Streptococcus gordonii DL1. Infect Immun. 2002;70(3):1209-1218. doi:10.1128/iai.70.3.1209-1218.2002
- 51. Kolenbrander PE, Andersen RN, Ganeshkumar N. Nucleotide sequence of the Streptococcus gordonii PK488 coaggregation adhesin gene, scaA, and ATPbinding cassette. Infect Immun. 1994;62(10):4469-4480. PMID: 7927711.
- 52. Klein RS, Harris CA, Small CB, Moll B, Lesser M, Friedland GH. Oral candidiasis in high-risk patients as the initial manifestation of the acquired immunodeficiency syndrome. N Engl J Med. 1984;311(6):354-358. doi:10.1056/nejm198408093110602

- 53. Metwalli KH, Khan SA, Krom BP, Jabra-Rizk MA. Streptococcus mutans, Candida albicans, and the human mouth: a sticky situation. Plos Pathog. 2013;9(10):e1003616. doi:10.1371/journal.ppat.1003616
- Klinke T, Guggenheim B, Klimm W, Thurnheer T. Dental caries in rats associated with Candida albicans. Caries Res. 2011;45(2):100-106. doi:10.1159/000324809
- 55. Xiao J, Moon Y, Li L, et al. Candida albicans Carriage in Children with Severe Early Childhood Caries (S-ECC) and Maternal Relatedness. Plos One. 2016;11(10):e0164242. doi:10.1371/journal.pone.0164242
- 56. Liu S, Qiu W, Zhang K, et al. Nicotine Enhances Interspecies Relationship between Streptococcus mutans and Candida albicans. Biomed Res Int. 2017;2017:7953920. doi:10.1155/2017/7953920
- 57. Falsetta ML, Klein MI, Colonne PM, et al. Symbiotic relationship between Streptococcus mutans and Candida albicans synergizes virulence of plaque biofilms in vivo. Infect Immun. 2014;82(5):1968-1981. doi:10.1128/iai.00087-14
- Raja M, Hannan A, Ali K. Association of oral candidal carriage with dental caries in children. Caries Res. 2010;44(3):272-276. doi:10.1159/000314675
- 59. Jarosz LM, Deng DM, van der Mei HC, Crielaard W, Krom BP. Streptococcus mutans competencestimulating peptide inhibits Candida albicans hypha formation. Eukaryot Cell. 2009;8(11):1658-1664. doi:10.1128/ec.00070-09
- 60. Heintze U. Secretion rate, buffer effect and number of lactobacilli and Streptococcus mutans of whole saliva of cigarette smokers and nonsmokers. Scand J Dent Res. 1984;92(4):294-301. doi:10.1111/j.1600-0722.1984.tb00894.x
- 61. Sakki T, Knuuttila M. Controlled study of the association of smoking with lactobacilli, mutans streptococci and yeasts in saliva. Eur J Oral Sci. 1996;104(5-6):619-622. doi:10.1111/j.1600-0722.1996.tb00151.x
- 62. Ravald N, Birkhed D, Hamp SE. Root caries susceptibility in periodontally treated patients. Results after 12 years. J Clin Periodontol. 1993;20(2):124-129. doi:10.1111/j.1600-051x.1993.tb00326.x
- 63. Wu J, Peters BA, Dominianni C, Zhang Y, Pei Z, Yang L et al. Cigarette smoking and the oral microbiome in a large study of American adults. ISME J. 2016;10(10):2435-2446. doi:10.1038/ismej.2016.37
- 64. Seneviratne CJ, Zhang CF, Samaranayake LP. Dental plaque biofilm in oral health and disease. Chin J Dent Res. 2011;14(2):87-94. https://cjdr.quintessenz.de/ index.php?doc=abstract&abstractID=22979. Accessed December 17, 2018.
- 65. Krzysciak W, Jurczak A, Koscielniak D, Bystrowska B, Skalniak A. The virulence of Streptococcus mutans and the ability to form biofilms. Eur J Clin Microbiol Infect Dis. 2014;33(4):499-515. doi:10.1007/s10096-013-1993-7
- 66. Macgregor ID, Edgar WM, Greenwood AR.

Effects of cigarette smoking on the rate of plaque formation. J Clin Periodontol. 1985;12(1):35-41. doi:10.1111/j.1600-051x.1985.tb01351.x

- 67. Huang R, Li M, Gregory RL. Bacterial interactions in dental biofilm. Virulence. 2011;2(5):435-444. doi:10.4161/viru.2.5.16140
- Vukosavljevic D, Custodio W, Buzalaf MA, Hara AT, Siqueira WL. Acquired pellicle as a modulator for dental erosion. Arch Oral Biol. 2014;59(6):631-638. doi:10.1016/j.archoralbio.2014.02.002
- 69. Evans P, Der G, Ford G, Hucklebridge F, Hunt K, Lambert S. Social class, sex, and age differences in mucosal immunity in a large community sample. Brain Behav Immun. 2000;14(1):41-48. doi:10.1006/brbi.1999.0571
- 70. Golpasand HL, Zakavi F, Ansarifar S, Ghasemzadeh O, Solgi G. Association of dental caries and salivary sIgA with tobacco smoking. Aust Dent J. 2013;58(2):219-223. doi:10.1111/adj.12059
- 71. Avsar A, Darka O, Bodrumlu EH, Bek Y. Evaluation of the relationship between passive smoking and salivary electrolytes, protein, secretory IgA, sialic acid and amylase in young children. Arch Oral Biol. 2009;54(5):457-463. doi:10.1016/j.archoralbio.2009.01.017
- 72. Struzycka I. The oral microbiome in dental caries. Pol J Microbiol. 2014;63(2):127-135. http://www.pjm. microbiology.pl/archive/vol6322014127.pdf. Accessed December 17, 2018.
- 73. Jette AM, Feldman HA, Tennstedt SL. Tobacco use: a modifiable risk factor for dental disease among the elderly. Am J Public Health. 1993;83(9):1271-1276. doi:10.2105/ajph.83.9.1271

ACKNOWLEDGEMENTS

We thank Xiaoge Jiang for her kind language editing.

CONFLICTS OF INTEREST

Authors have completed and submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest and none was reported.

FUNDING

This work was partially supported by the National Natural Science Foundation of China (81400501 to ML and 31800114 to RH).

PROVENANCE AND PEER REVIEW

Not commissioned; externally peer reviewed.